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Title of the paper: Genetic markers in a multi-ethnic sample for childhood acute lymphoblastic leukemia risk

Running title: Genetic markers for childhood leukemia

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ABSTRACT

Genome-wide association studies have identified multiple risk loci for childhood acute lymphoblastic leukemia (ALL), but mostly in European/White populations despite Hispanics having a greater risk. We re-examined SNPs of known associations with childhood ALL and known HLA region lymphoma risk markers in a multi-ethnic population. Significant associations were found in two ARID5B variants (rs7089424 and rs10821936). We replicated a strong risk association in non-Hispanic White males with rs2395185, a protective marker for lymphoma. Another HLA region marker, rs2647012, showed a risk association among Hispanics only, while a strong protective association was found with rs1048456, a follicular lymphoma risk marker. Our study validated this new case-control sample by confirming genetic markers associated with childhood ALL, and yielded new associations with lymphoma markers. Despite positive results, our study did not provide any clues to why Hispanics have a higher susceptibility to childhood leukemia, suggesting that environmental factors may have a strong contribution.
INTRODUCTION

Over a third of cancers in children are leukemias [1], with acute lymphoblastic leukemia (ALL) being the most common [2-4]. Approximately 3.8 individuals (less than 14 years of age) per 1,000,000 are diagnosed with childhood ALL per year in the United States [5]. The racial disparity in the incidence of childhood ALL has been well established [6,7]. In the United States, incidence of ALL is greatest in Hispanic children, followed by non-Hispanic Whites, Asians, and then Blacks [7-9].

Research continually tries to determine the etiology of childhood leukemias. While there are a few known risk factors associated with childhood ALL [10-15], more than 90% are of unknown etiology [11]. Increased birth weight is an established risk factor shown to increase risk of childhood ALL [16-18]. Environmental factors are also involved, and may work in conjunction with genetic factors to cause many cancers [10].

Genome-wide association studies (GWAS) [19-22] have identified multiple risk loci showing significant associations with childhood ALL. Most significantly, variants located within the ARID5B, IKZF1, and CEBPE genes have robust risk associations. Most of these studies, however, have only evaluated risk among those of European ancestry. Two multiethnic studies conducted by Xu et al. looked at various risk loci in both African American and Hispanic American populations [8,22], discovering that some markers are universal across races/ethnicities, while others are race/ethnic-specific.

We hypothesized that the variation in ethnic/racial susceptibility to childhood ALL has a genetic basis. We re-examined five previously discovered single nucleotide polymorphisms (SNPs) of known associations to leukemia (rs7089424, rs10821936, rs10994982, rs4132601, and rs2239633), along with three HLA region susceptibility markers for lymphomas, since
lymphoma and ALL both stem from lymphoid cells. These included rs2395185, a marker of HLA DRB4 lineage [23] which has previously shown associations with major leukemias including childhood ALL [24,25] and other diseases including Hodgkin lymphoma [26], lung cancer [27], rheumatoid arthritis [28], asthma [29], and ulcerative colitis [30-32]; rs10484561, which has been shown to be a strong risk marker in follicular lymphoma (FL) [33]; and rs2647012, which is a protective marker for FL [34]. These SNPs were examined in a multi-ethnic sample (non-Hispanic Whites, Hispanic Whites, and Blacks) from Houston, Texas to assess their association with ALL.

SUBJECTS AND METHODS

Study population

Institutional Review Board (IRB) approval was obtained at both the Baylor College of Medicine (BCM) and Florida International University prior to the start of the study. The case-control study was comprised of 161 incident childhood ALL cases and 231 healthy frequency-matched controls for gender, contemporaneously recruited at the Texas Children’s Cancer Center in Houston, TX from 2007 to 2012. The children were less than 18 years of age at diagnosis, and exclusion criteria for both cases and controls were refusal to participate in the study and the diagnosis of any other disease or cancer. Subjects or their parents provided informed consent for provision of epidemiological data with a questionnaire and a biological sample. The DNA samples were extracted from saliva or peripheral blood samples at BCM. Race/ethnicity was determined by the responses provided on the questionnaire. Parents were requested to state the race (White, Black/African American, Asian, American Indian/Alaska Native, or Native Hawaiian/Other Pacific Islander) and ethnicity (Hispanic/Latino or non-Hispanic/Latino) of the
child. The questionnaire also collected the race/ethnicity of the parents, which was used to verify the response. Information on clinical subtype of ALL was collected from medical records.

Genotyping

The main features for the SNPs genotyped are shown in Table I. Pre-developed TaqMan® SNP Genotyping Assays (LifeTech, Foster City, CA) were used for all of the SNPs we examined. Genotyping was achieved using the Bio-Rad CFX96 real-time PCR machine (Hercules, CA). The TaqMan assays consist of singleplex reactions carried out in ninety-six well plates. Each plate contained two no template controls (NTCs), a positive control, and random replicate samples. Bio-Rad SsoFast™ Probes Supermix, a 2x reaction buffer which contains the necessary components for running the PCR; Sso7d-fusion polymerase, dNTPs, MgCl₂, and stabilizers, was used with the TaqMan Assay. PCR amplifications were performed using the manufacturer’s suggestion of 20 μL total volume and with the following PCR thermal cycling conditions: enzyme activation at 95°C for two minutes, and 49 cycles of denaturation at 95°C for 5 seconds followed by annealing and extension at 61°C for 5 seconds. Bio-Rad CFX Manager software (version 3.0) was used for data acquisition and genotype assignment.

To adjust for heterogeneity in our sample, especially in Hispanics, we used two ancestry-informative markers, AIMs, to control for potential confounding caused by population stratification and to avoid spurious associations [35]. The two AIMs we used, rs285 and rs2891, have been previously used in Hispanic populations to account for the differences in genetic ancestry [36,37]. The risk associations were adjusted by each of the AIMs besides adjustment by self-declared race/ethnicity.

Statistical analysis
Statistical analyses were performed using Stata v.11 (StataCorp, College Station, TX). Pearson’s $X^2$, Student's t-test (for means) or median test (for medians) were used to compare characteristics between the cases and controls. Logistic regression methods were used to calculate crude and adjusted odds ratios (OR) and 95% confidence intervals (CIs). All statistical tests were two-tailed, and the threshold for statistical significance was set at $P \leq 0.05$. The ORs, with 95% CIs, were used as a measure of effect size. Genotype counts were tested for Hardy–Weinberg equilibrium (HWE) in controls for each SNP. By default, we used the additive genetic model to assess associations by Cochrane-Armitage trend test. Due to its previous association being in the recessive model, rs2395185 was analyzed also for the recessive model association. Ethnic- and gender-specific associations were calculated through stratified analyses after grouping subjects as non-Hispanic White, Hispanic White, Blacks, and Others. Associations in the overall sample were assessed by adjustment for race/ethnicity as well as by Mantel-Haenszel analysis after stratification. None of the SNPs were located in coding regions, therefore we used RegulomeDB (Stanford University, Palo Alto, CA) to determine their scores for regulatory effects.

**RESULTS**

All cases and controls were genotyped for the eight candidate SNPs and two ancestry-informative markers (AIMs). Genotype call rates were greater than 95% for both cases and controls. Table II shows characteristics of the case-control sample. The case samples included 86 males (53%) and 75 females (47%). Out of these cases, 66 identified themselves as non-Hispanic White, 72 as Hispanic White, 17 as Black, and 6 as “other.” The group labeled “other” included those identifying themselves as Asian, Native American, or other. The healthy controls included 130 males (56%) and 101 females (44%), who had visited a pediatric clinic at Texas
Children's Hospital for a non-disease related reason. Forty-nine were classified as non-Hispanic White, 98 as Hispanic White, and 78 as Black. The distribution of ethnic background was different between cases and controls mainly due to the infrequency of childhood ALL in Blacks. Because of this difference, results were adjusted for ethnic background or stratified analyses were performed when necessary. Mean birth weight was not different between cases and controls. Eighty-eight percent of the cases were diagnosed with early precursor B (early pre-B) ALL subtype, and associations did not change in effect size depending on the molecular subtype. There were no significant genotype associations found within the Black subpopulation of the sample and each of the SNPs genotyped (results not shown), possibly due to the small number of cases in the population sample.

**GWAS risk markers**

Results including genotype frequencies and ORs are described in Tables III-IV. Analyses yielded significant associations with some of the genetic markers similar to previous reports. In total, three *ARID5B* SNPs were examined for associations. Two of the *ARID5B* SNPs, rs7089424 and rs10821936, showed expected risk associations, while rs10994982 did not show an association (Table III). The SNP rs7089424 had an overall OR per allele (OR<sub>allele</sub>) of 1.69 (**<sub>0.001**). The association showed a somewhat stronger risk in the non-Hispanic subgroup (OR<sub>allele</sub>=2.11, **<sub>0.01**), compared with Hispanics (OR<sub>allele</sub>=1.61, **<sub>0.02**). Similarly, rs10821936 had an overall OR<sub>allele</sub>=1.48 (**<sub>0.05**). Adjustments of the analyses for race/ethnicity did not change the observed results. Adjustment for AIMs did not appreciably alter the results either.

Hardy-Weinberg equilibrium was violated in controls for *IKZF1* rs4132601, which could lead to spurious results, and was therefore excluded. There was no significant association found for *IKZF1* rs4132601 or for *CEBPE* rs2239633.
HLA region lymphoma risk markers

The SNP located in the HLA-DR region, rs2395185, showed a weak, non-significant risk overall (Table IV). This SNP is an exclusive marker for the HLA-DRB4 (DR53) lineage [23]. Since this lineage was shown to be a risk marker for childhood ALL in European samples, but only in males [24], we examined rs2395185 association in males. The non-Hispanic White male group had an OR_{allele} of 2.79 ($P=0.016$). The OR reached 6.21 (95% CI=0.70-54.96) for homoyzgosity for the variant allele, which corresponds to the original association [24]. The known protective marker for follicular lymphoma, rs2647012, showed a statistically significant association in Hispanics OR_{allele} =2.21 ($P=0.007$), but not in non-Hispanics ($P_{interaction}= 0.003$ for ethnicity). The significance remained after adjusting for both rs23951885 and rs10484561. The variant rs10484561 was shown to be a strong protective marker in this study, opposite of what was found in follicular lymphoma [33]. Using the recessive model, the variant allele showed a strong association, in the overall sample with OR_{rec}=0.19 ($P=0.009$) after adjustment for race/ethnicity, and OR_{rec}=0.17 ($P=0.004$) in Mantel-Haenszel analysis.

DISCUSSION

Despite being well established, the racial disparity in the incidence of childhood ALL is not always addressed in genetic association studies. Most GWAS, until recently, have identified risk loci using only European-origin populations. Variant polymorphisms located within the ARID5B, IKZF1, and CEBPE genes have reported strong risk associations in multiple studies [19-22, 38-40]. Our study provides some confirmation of previously discovered genetic markers associated with childhood ALL, which also validated our case-control set for further exploration.

Of the three ARID5B SNPs, rs7089424 and rs10821936 showed significant risk associations. The ARID5B gene is involved in transcriptional regulation during embryonic
Overexpression of the gene in particular leukemias have led some to speculate that variations within the gene may affect B-lineage development, and increase susceptibility to B-lineage leukemia [20]. The marker rs7089424 was associated with a stronger risk of leukemia in the non-Hispanic subgroup compared with Hispanics, replicating recent findings from case-control studies using Hispanic populations [8,22].

In the recent multi-ethnic GWAS by Xu et al., rs10821936 was found to be a significant risk marker across all ethnicities. Xu et al. noted that the risk allele frequencies for rs10821936 increased in order by race incident rates: Black/African Americans, non-Hispanic/European American, and Hispanic Americans [22]. Our results showed a similar trend with an increasing risk allele frequency in cases of Blacks, non-Hispanic Whites and Hispanics. The multi-ethnic GWAS reported that rs10821936 was highly correlated with Native American genetic ancestry [22], substantiating their previous observations that Native American ancestry correlates with higher risk of relapse in Hispanics, and leading to speculation that this may be a factor with the increased risk of childhood ALL for Hispanic children who have a high proportion of Native American ancestry [41].

The SNP rs4132601, located in the Ikaros family zinc finger 1 (IKZF1) gene, is associated with increased risk of childhood ALL in multiple studies [19,21,39,40,42]. The Ikaros proteins are known to be involved with lymphocyte development and differentiation [19], and deletions are frequent and associated with unfavorable prognosis in B-cell precursor ALL [19,43]. One study found this variant to be a significant risk marker amongst non-Hispanic Whites, but not in Hispanics, despite similar allele frequencies [40]. Chokkalingam et al. hypothesized that that this marker’s association may be due to linkage disequilibrium with a functional variant, and because of admixture in Hispanic populations the linkage disequilibrium
may vary [40]. Our study was unable to examine this SNP, due to Hardy-Weinberg
disequilibrium found in controls after stratification for race/ethnicity.

The present study confirmed some, but not all previous findings of GWAS. With the
\textit{ARID5B} risk SNPs, there was heterogeneity even between the first two GWAS reports [19,20].
The modest sample size we had also reduced the statistical power of our study. Nevertheless,
confirmed results validated the present case-control sample for further genetic association
studies.

The relevance of lymphoma-associated polymorphisms in childhood ALL was assessed
by genotyping rs2395185, rs10484561, and rs2647012. The SNP near the \textit{HLA-DRA} gene,
rs2395185, is a marker for the \textit{HLA-DRB4} (DR53) lineage [23]. The \textit{HLA-DRB4} lineage or its
marker SNP have been previously shown as a risk marker in lung cancer [27], asthma [29],
rheumatoid arthritis [28], type I diabetes [44], adult acute myeloblastic leukemia [45], chronic
myeloid leukemia [46], chronic lymphoid leukemia [47-49] and in childhood ALL (males
only)[24,50,51], and as a protective marker for non-Hodgkin lymphoma [26], and ulcerative
colitis [30-32,52]. The DRB4/DR53 lineage has been shown previously to have a risk association
with childhood ALL, with male specificity, within a European sample via HLA typing [24]. The
first GWAS association of rs2395185 was with ulcerative colitis [30,32]. The variant allele, T,
was later found to be a protective marker in a GWAS examining risk factors for classical
Hodgkin lymphoma [26], and most recently a risk marker factor in Asian females for lung cancer
[27]. Our results replicated the strong male specificity of the risk for childhood ALL, specifically
in non-Hispanic White males, with no association in Hispanics. The DRB4 lineage has unique
features, such as lower expression levels of HLA-DR molecules, poor interaction with CD4,
disrupted intracellular transport, and possibly contains extra amount of DNA in the DR/DQ
region which may contribute to this risk association in childhood ALL [25]. One important finding of the present study is that the risk modifiers of lymphoma showed associations in opposite directions in childhood ALL.

The variant rs2647012, a marker for DRB3/DRB5 lineages [53] and a protective marker for lymphoma [34], showed a statistically significant risk association among Hispanics, but not in non-Hispanics ($P_{\text{interaction}} = 0.003$ for ethnicity). Our study appears to be the first looking at a Hispanic population with this SNP, and further studies are warranted to determine if this inverse relationship of risk exists in non-Hispanic and Hispanic Whites. Variant rs10484561 showed signs of being a protective marker for childhood ALL, opposite to the follicular lymphoma findings [33].

It is now customary that the additive model is used to assess statistical associations of SNPs. While the additive model has sufficient power to detect associations in most situations, there are certain scenarios that it may not show statistical significance when in fact, there is an association. An association conforming to the strictly recessive model when allele frequency is low is one example [54-56]. For this reason, and also because the original rs2395185 association was a recessive model association, we also assessed this genetic model in HLA region associations. This approach consistently yielded larger effect sizes for HLA region SNPs, especially for rs2395185 as in previous studies. We are in favor of routine use of the recessive model analysis in exploration of associations in the HLA region.

Our study had a number of limitations. With childhood ALL being a rare disease, the sample size for our study was small, resulting in limited statistical power. The issue of self-reported ethnicity may be of concern. This method is common with population-based association studies, and residual confounding is often suspected. Even though it has been described that self-
reported ethnicity may be reliable [57], the heterogeneity within the Hispanic population is still a concern. A recent study conducted in a Spanish population was unable to replicate original risk associations found in Hispanic Americans, demonstrating the large heterogeneity in this high risk group [22,58]. To adjust for heterogeneity in our sample, especially in Hispanics, we used two ancestry-informative markers, AIMs, to control for confounding caused by population stratification and to avoid spurious associations [35]. The AIMs have widely different allele frequencies in major human continental groups. The two AIMs we used, rs285 and rs2891, have been previously used in Hispanic populations to account for the differences in genetic ancestry [36,37]. Adjusting the risk associations by AIMs did not alter the results.

Our study did have a well-defined phenotype, with molecular ALL types determined. The use of a multi-ethnic sample population was a strength, especially for the ethnic disparity that exists in childhood ALL susceptibility. Another strength is the use of multiple genetic models, where appropriate, to determine associations that may remain undetectable by the exclusive use of the additive model association. The replication of known leukemia markers validated our sample set for further studies. This pilot study is part of an ongoing effort at BCM. Recruitment for the second phase is continuing together with clinical follow-up.

In summary, we validated a new multi-ethnic case-control set and also examined some new markers with their association with childhood ALL. The examination of lymphoma risk markers yielded associations in opposite directions for childhood ALL, and also confirmed a previously identified childhood ALL risk marker. Two HLA region associations were ethnicity-specific. Still, our study did not provide clues as to why Hispanics have a higher susceptibility to childhood ALL, suggesting that environmental factors may have to be incorporated in future studies to examine their role in this differential. Studies with information on environmental
exposures may help explain how gene-environment interactions contribute to childhood ALL susceptibility and its variation among different populations.
ACKNOWLEDGMENTS

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POTENTIAL CONFLICTS OF INTERESTS

Disclosure forms provided by the authors are available with the full text of this article at www.informahealthcare.com/lal.
REFERENCES


Table I. Main features of SNPs analyzed

<table>
<thead>
<tr>
<th>Gene</th>
<th>SNP</th>
<th>Chromosome nucleotide position</th>
<th>Inclusion criteria</th>
<th>PMID**</th>
<th>Minor allele and frequency†</th>
<th>Location</th>
<th>RegulomeDB score††</th>
</tr>
</thead>
<tbody>
<tr>
<td>ARID5B</td>
<td>rs7089424</td>
<td>chr10: 63752159</td>
<td>GWAS identified risk loci for childhood ALL</td>
<td>19684604, 22660188, 20042726</td>
<td>(G) 0.314</td>
<td>Intrinsic</td>
<td>3a</td>
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<td>ARID5B</td>
<td>rs10821936</td>
<td>chr10: 63723577</td>
<td>GWAS identified risk loci for childhood ALL</td>
<td>19684603, 20054350, 22660188, 23512250, 22291082</td>
<td>(C) 0.318</td>
<td>Intrinsic</td>
<td>5</td>
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<td>rs10994982</td>
<td>chr10: 63710104</td>
<td>GWAS identified risk loci for childhood ALL</td>
<td>19684603, 22660188</td>
<td>(A) 0.457</td>
<td>Intrinsic</td>
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<td>IKZF1</td>
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<td>19684604, 22660188, 20054350</td>
<td>(G) 0.306</td>
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<td>CEBPE</td>
<td>rs2239633</td>
<td>chr14: 23589057</td>
<td>GWAS identified risk loci for childhood ALL</td>
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<td>(A) 0.466</td>
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<td>HLA-DR region</td>
<td>rs2395185</td>
<td>chr6: 32433167</td>
<td>HLA-DRB4/DR53 lineage; ALL risk marker (in European males); Hodgkin lymphoma risk marker</td>
<td>10397736, 12008082, 22286212, 7909466</td>
<td>(T) 0.423</td>
<td>Intrinsic</td>
<td>6</td>
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<tr>
<td>HLA-DQB1 region</td>
<td>rs2647012</td>
<td>chr6: 32664458</td>
<td>HLA-DRB3/DRB5 lineage, protective marker for non-Hodgkin (follicular) lymphoma</td>
<td>21533074, 22911334, 23455380</td>
<td>(T) 0.381</td>
<td>Intergenic</td>
<td>6</td>
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<tr>
<td>HLA-DQA1 region</td>
<td>rs10484561</td>
<td>chr6: 32665420</td>
<td>HLA-DR1/DR10 lineage, risk marker in follicular lymphoma</td>
<td>20639881, 21533074, 23025665</td>
<td>(G) 0.084</td>
<td>Intergenic</td>
<td>6</td>
</tr>
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</table>


**PubMed identifier number

†Minor allele frequencies are from a reference Caucasian population (U.S. residents of northern and western European ancestry) genotyped in HapMap project

††RegulomeDB scores range from 1 (most functional) to 5 (least functional) (6=other). Not all SNPs have a RegulomeDB score [http://regulome.stanford.edu/]
Table II. Characteristics of cases and controls

<table>
<thead>
<tr>
<th></th>
<th>Cases n=161</th>
<th>Controls n=231</th>
<th>P value</th>
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</thead>
<tbody>
<tr>
<td><strong>Ethnic background</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Non-Hispanic White</td>
<td>66</td>
<td>49</td>
<td>&lt;0.001</td>
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<td>Hispanic White</td>
<td>72</td>
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</tr>
<tr>
<td>Black</td>
<td>17</td>
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<td></td>
</tr>
<tr>
<td>Other*</td>
<td>6</td>
<td>6</td>
<td></td>
</tr>
<tr>
<td><strong>Gender</strong></td>
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<tr>
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<tr>
<td>Ratio</td>
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<td>1.29</td>
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<tr>
<td><strong>Birth weight (grams)</strong></td>
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</tr>
<tr>
<td>Mean (SD)</td>
<td>3349.3 (584)</td>
<td>3263.3 (684)</td>
<td>0.23</td>
</tr>
<tr>
<td>Median (IQR)</td>
<td>3400 (760)</td>
<td>3311.5 (850)</td>
<td>0.48</td>
</tr>
</tbody>
</table>

*Other includes Asian, Native American, or any other ethnicities
Table III. SNP associations previously shown as ALL risk markers in GWAS (overall*)

<table>
<thead>
<tr>
<th>Gene</th>
<th>SNP</th>
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<th>A/A</th>
<th>A/B</th>
<th>B/B</th>
<th>A/A</th>
<th>A/B</th>
<th>B/B</th>
<th>Cases</th>
<th>Controls</th>
<th>OR&lt;sub&gt;allele&lt;/sub&gt; **(95% CI)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>ARID5B</td>
<td>rs7089424</td>
<td>G</td>
<td>37</td>
<td>80</td>
<td>43</td>
<td>113</td>
<td>68</td>
<td>46</td>
<td>0.52</td>
<td>0.35</td>
<td>1.69 (1.28-2.24)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>ARID5B</td>
<td>rs10821936</td>
<td>C</td>
<td>39</td>
<td>72</td>
<td>44</td>
<td>104</td>
<td>69</td>
<td>50</td>
<td>0.52</td>
<td>0.38</td>
<td>1.48 (1.12-1.95)</td>
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<td>rs10994982</td>
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<td>78</td>
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<td>90</td>
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<td>43</td>
<td>0.41</td>
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<td>1.00 (0.75-1.34)</td>
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<td>IKZF1</td>
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<td>16</td>
<td>141</td>
<td>56</td>
<td>29</td>
<td>0.31</td>
<td>0.25</td>
<td>1.19 (0.88-1.60)</td>
<td>0.269†</td>
</tr>
<tr>
<td>CEBPE</td>
<td>rs2239633</td>
<td>A</td>
<td>68</td>
<td>68</td>
<td>23</td>
<td>118</td>
<td>74</td>
<td>36</td>
<td>0.36</td>
<td>0.32</td>
<td>1.02 (0.76-1.36)</td>
<td>0.881</td>
</tr>
</tbody>
</table>

*ORs adjusted for self-reported race/ethnicity
**OR per allele (OR<sub>allele</sub>) for the additive model
†Hardy-Weinberg disequilibrium in controls
Table IV. Association of HLA region lymphoma susceptibility markers (overall*)

<table>
<thead>
<tr>
<th>SNP</th>
<th>Gene</th>
<th>Minor allele</th>
<th>Cases</th>
<th>Controls</th>
<th>Minor allele frequency</th>
<th>OR_{allele}** (95% CI)</th>
<th>P value</th>
<th>OR_{rec}† (95% CI)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs2395185</td>
<td>HLA-DR region</td>
<td>T</td>
<td>69</td>
<td>68</td>
<td>22</td>
<td>0.35</td>
<td>0.28</td>
<td>1.27 (0.94-1.71)</td>
<td>0.127</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>120</td>
<td>72</td>
<td>25</td>
<td></td>
<td></td>
<td>1.1 (0.65-2.26)†‡</td>
<td>0.553</td>
</tr>
<tr>
<td>rs2647012</td>
<td>HLA-DQB1 region</td>
<td>A</td>
<td>78</td>
<td>64</td>
<td>16</td>
<td>0.3</td>
<td>0.28</td>
<td>1.09 (0.80-1.47)</td>
<td>0.595</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>116</td>
<td>47</td>
<td>30</td>
<td></td>
<td></td>
<td>0.57 (0.30-1.12)</td>
<td>0.103</td>
</tr>
<tr>
<td>rs10484561</td>
<td>HLA-DQA1 region</td>
<td>G</td>
<td>128</td>
<td>23</td>
<td>3</td>
<td>0.09</td>
<td>0.15</td>
<td>0.70 (0.46-1.06)</td>
<td>0.094</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>149</td>
<td>21</td>
<td>18</td>
<td></td>
<td></td>
<td>0.19 (0.05-0.66)</td>
<td>0.009</td>
</tr>
</tbody>
</table>

*ORs adjusted for self-reported ethnicity and race
**OR per allele (OR_{allele}) for the additive model
†OR recessive (OR_{rec}) for the variant homozygous genotype
‡OR_{allele}=1.88 (P=0.003) in males; OR_{allele}=2.79 (P=0.016) in non-Hispanic White males